

Anisakidae IgG ELISA

Enzyme immunoassay for the diagnosis of human digestive anisakidosis

96 assays on individual wells for in vitro diagnostic use and for professional laboratory use



Instructions for use for article N° 9800
UDI-DI: 07640158219805



Intended use:

The Bordier *Anisakidae* IgG ELISA kit is intended for the qualitative detection of IgG antibodies against parasites of *Anisakidae* family in human serum. Serology is an aid for diagnosis and cannot be used as the sole method of diagnosis.

Background:

Anisakidosis is a worldwide helminthic zoonosis caused by some sea fish nematodes of the family *Anisakidae*. Humans can be infected by ingesting raw or undercooked contaminated fish meat. Adult worms of anisakid nematodes reside in the digestive tract of marine mammals (final hosts). After being embryonated in the water, the eggs release infesting L3 larvae into the water, which are ingested by the planktonic crustacean (intermediate host). After consumption by fish or cephalopods (paratenic hosts), the larvae migrate into the viscera and the peritoneal cavity so that they no longer evolve. After ingestion by marine mammals, the larvae develop into adult worms. Man, accidental host, is a parasitic dead end. Most infected people do not show any symptoms. However, in some cases, symptoms appear at the gastric stage (epigastric pain, nausea and vomiting), intestinal (abdominal pain, nausea, vomiting and diarrhea) or at the allergic stage (hives, pruritus, angioedema and bronchospasm). Diagnosis is based on signs and symptoms plus a history of exposure and a positive result by serological testing.

Principle and presentation:

The kit provides all the material needed to perform 96 enzyme-linked immunosorbent assays (ELISA) on breakable microtitration wells sensitized with *Anisakidae* excreted/secreted (E/S) larval antigens. Specific antibodies in the sample will bind to these antigens and washing will remove unspecific antibodies. The presence of parasite specific antibodies is detected with a Protein A - alkaline phosphatase conjugate. A second washing step will remove unbound conjugate. Revealing bound antibodies is made by the addition of pNPP substrate which turns yellow in the presence of alkaline phosphatase. Color intensity is proportional to the amount of *Anisakidae* specific antibodies in the sample. Potassium phosphate is added to stop the reaction. Absorbance at 405 nm is read using an ELISA microplate reader.

The test is manual but can be performed with automatic systems, which must be validated by the user.

Material contained in the kit (96 assays):

WELL	9800-01	Breakable ELISA strips sensitized with <i>Anisakidae</i> E/S antigens	96	wells
DILB	9800-02	Dilution buffer (10 x) concentrate, coloured purple	50	ml
WASH	9800-03	Washing solution (10 x) concentrate	50	ml
ENZB	9800-04	Enzyme buffer	50	ml
STOP	9800-05	Stopping solution (0.5M K ₃ PO ₄)	25	ml
CONTROL -	9800-06	Negative control serum (20 x), green cap	200	µl
CONTROL -/+	9800-07	Weak positive control serum (cut off, 20 x), yellow cap	200	µl
CONTROL +	9800-08	Positive control serum (20 x), red cap	200	µl
CONJ	9800-09	Protein A - alkaline phosphatase conjugate (50 x), purple cap	300	µl
SUBS	9800-10	Phosphatase substrate (para-nitrophenylphosphate)	20	tablets
		Multipipette reservoir, 25 ml	1	piece
		Frame for ELISA 8-well holder	1	piece

Shelf life and storage:

Store the kit between +2°C and +8°C (transport validated between -20°C and +37°C for 21 days), avoid long term exposure of the components to direct light. The expiry date and the lot number of the kit are printed on the side of the box. After initial opening, all reagents are stable until the expiry date when stored between +2°C and +8°C.

Equipment needed but not provided with the kit:

Pipettes (ml and µl). Flasks. Dilution tubes. Adhesive tape to cover wells during incubations. Distilled water. Incubator set at +37°C. ELISA reader set at 405 nm. Manual or automatic equipment for rinsing wells. Vortex mixer. Timer.

Preparation of reagents before use:

Bring all reagents between +20°C and +30°C for 1 hour minimum and mix before use.

ELISA wells: open side of aluminium bag 9800-01 and remove number of wells needed (one for blank, three for controls plus the number of samples). Place sensitized wells in 8-well holder(s). If needed, complete the empty positions in the holder with used wells. Insert holder(s) in the frame in the correct orientation. Reseal open package with desiccant pad.

Dilution buffer: dilute dilution buffer (10 x) concentrate 9800-02, 1/10 in distilled water. This is used for the dilution of controls, samples, and conjugate. The diluted buffer is stable for 2 months between +2°C and +8°C.

Washing solution: dilute washing solution (10 x) concentrate 9800-03, 1/10 in distilled water. You may also use your own washing solution. Avoid buffers containing phosphate which could inhibit the enzymatic activity of the alkaline phosphatase. The diluted washing solution is stable for 2 months between +2°C and +8°C.

Control sera: dilute 10 µl control sera 9800-06 to -08 in 190 µl dilution buffer solution (final dilution 1/20). The diluted control sera are stable for 2 months between +2°C and +8°C.

Conjugate: dilute conjugate 9800-09 in dilution buffer solution (final dilution 1/50). Dilute conjugate on the day of the assay. Do not store diluted conjugate.

Substrate solution: dissolve tablet(s) of phosphatase substrate 9800-10 in undiluted enzyme buffer 9800-04 (1 tablet in 2.5 ml buffer). Vortex until complete dissolution of the tablet(s). Dilute substrate on the day of the assay and protect the tube from direct light. Tablets and substrate solutions should be colourless or should have only a slight yellow tinge. If a tablet or a substrate solution turns yellow, it may have been partially hydrolysed and should be discarded. Do not store the substrate solution.

Stopping solution: use reagent 9800-05 undiluted.

Specimen collection and preparation:

Use human serum. Store between +2°C and +8°C if analyzed within 7 days, otherwise store at -20°C or lower. Avoid repeated freezing and thawing. Vortex samples and dilute 1/201 in dilution buffer solution (for instance 5 µl sample in 1.0 ml). Do not store diluted samples.

Warnings and precautions:

Toxic compounds are found in following concentration:

Component	Reference	Sodium azide (NaN ₃)	Merthiolate
Dilution buffer (10 x)	9800-02	0.1 %	0.02 %
Washing solution (10 x)	9800-03	0.05 %	/
Enzyme buffer	9800-04	0.01 %	/
Control sera (20 x)	9800-06 to -08	0.1 %	0.02 %
Conjugate (50 x)	9800-09	0.1 %	/

At the used concentrations, sodium azide and merthiolate do not have any toxicological risk on contact with skin and mucous membranes.

Component	Dangerous component	Danger pictogram	Hazard statement	Precautionary statement
Stopping solution	Potassium phosphate, tribasic		Causes serious eye damage	Wear eye protection. IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing.

- The negative, cut off and positive control sera (9800-06 to -08) are of animal origin (rabbits) and must be handled with care.
- Treat all reagents and samples as potentially infectious material.
- Do not interchange reagents of different lots or Bordier ELISA kits.
- Do not use reagents from other manufacturers with reagents of this kit.
- Do not use reagents after their expiry date.
- Close reagent vials tightly immediately after use and do not interchange screw caps to avoid contamination.
- Use separate and clean pipettes tips for each sample.
- Do not re-use microwells.
- Avoid deterioration of the microwells by mechanical action (tips/cones, nozzles).
- The descriptions of symbols used on the labels can be found on the website www.bordier.ch.

Disposal consideration:

All materials used for this test are generally considered as hazardous waste. Refer to national and regional laws and regulations for the disposal of hazardous waste.

Procedure:

When running the assay, avoid the formation of bubbles in the wells.

Step 1: Preincubation:

Fill wells with 250 µl of dilution buffer solution.

Incubate for 5 to 15 minutes between +20°C and +30°C.

Remove dilution buffer by aspiration or by shaking the wells over the sink.

Step 2: Incubation with samples:

Fill the first well of the first strip with 100 µl dilution buffer only (no-serum blank).

Fill the subsequent three wells with respectively 100 µl diluted negative, weak positive (cut off) and positive control serum. For assays of more than 25 samples, we recommend filling the three last wells with control sera as a duplicate.

Fill remaining wells with the diluted samples (100 µl each).

Cover wells with adhesive tape and incubate for 30 minutes at +37°C.

Remove sera and wash 4 x with ~ 250 µl washing solution.

Step 3: Incubation with conjugate:

Distribute 100 µl diluted conjugate in each well (including no-serum blank).

Cover wells with adhesive tape and incubate for 30 minutes at +37°C.

Remove conjugate and wash 4 x with ~ 250 µl washing solution.

Step 4: Incubation with substrate:

Distribute 100 µl substrate solution per well.

Cover wells with adhesive tape and incubate for 30 minutes at +37°C.

Stop the reaction by the addition of 100 µl stopping solution to each well.

Step 5: Measurement of absorbances:

If needed, wipe the bottom of the wells and eliminate bubbles. Measure absorbances at 405 nm within 1 hour after the addition of stopping solution.

Interpretation:

Subtract the value of the no-serum blank from all measured values. When applicable calculate the mean absorbance values of duplicated serum controls. The test is valid if the following criteria are met:

- Absorbance (A) of positive control > 1.200
- A of weak positive control > 9 % of A of positive control
- A of negative control < 8 % of A of positive control
- A of no-serum blank < 0.350

In case of sample providing signal overflowing the microplate reader measurement range, the value corresponding to the upper measurement range of the reader should be attributed.

Quality controls of current lots are published on our website: www.bordier.ch.

The antibody concentration of the weak positive (cut off) serum 9800-07 has been set to discriminate optimally between sera of clinically documented cases of anisakidosis and healthy human sera.
The cut off index of a sample is defined, after subtraction of the no-serum blank, as:

$$\text{Index} = \frac{\text{Absorbance sample}}{\text{Absorbance cut off serum}}$$

The result is **negative** when the index of the analyzed sample is lower than **1.0**. In this case, the IgG antibody concentration against **Anisakidae** antigens is clinically non-significant.

The result is **positive** when the index of the analyzed sample is higher or equal to **1.0**. In this case, the IgG antibody concentration against **Anisakidae** antigens is considered as clinically significant. It indicates that the patient has had a contact with the parasite.

A grey zone could be defined by each laboratory according to its patients population. In case of borderline or doubtful results, we recommend repeating the test again 2-4 weeks later with a fresh sample.

In case of positive or doubtful result, we recommend performing a confirmation test (most often by western blot) if such a test is available or required by national regulations.

Analytical performances:

Analytical specificity:

A specificity of 81% was found with 47 sera of patients with other parasitic infections. Cross-reactivity mainly occur in patients with toxocarosis, filarioses and strongyloidosis.

No positive or negative interference was observed with supraphysiological concentrations of hemoglobin, lipids or bilirubin in sera supplemented with interferents.

Precision:

Repeatability was assessed by testing 2 human serum samples in 24 wells on 1 assay.

Reproducibility was assessed by testing the 2 human serum samples in duplicate on 10 different assays.

	Repeatability		Reproducibility	
	Sample 1	Sample 2	Sample 1	Sample 2
Average (absorbance)	0.108	1.584	0.121	1.754
Standard deviation (absorbance)	0.008	0.082	0.020	0.111
Variation coefficient (%)	7.4	5.2	16.7	6.3

Clinical performances:

Diagnostic sensitivity:

A sensitivity of 97% was found with 38 sera of patients suffering from digestive and/or allergic anisakidosis.

Diagnostic specificity:

A specificity of 97 % was found with 180 sera of blood donors (Swiss). A specificity of 96% was found with 98 sera from patients of an infectiology unit (Swiss). A specificity of 86% was found with 43 sera from anisakidosis suspected patients, but where this disease has been certainly ruled out.

Positive and negative predictive value:

A PPV of 77% and a NPV of 99% were found with the populations mentioned above.

Expected values in normal and affected populations:

In a normal population of 180 Swiss blood donors and 98 sera from a Swiss infectiology unit, the expected Index value is 0.34. In an affected population of 47 sera from patients suffering from anisakidosis, the expected Index value is 3.71.

Incidents:

Any serious incident occurring in connection with the device shall be notified to the manufacturer and to the competent authority of the Member State in which the user and/or the patient is established.

Limitations:

Diagnosis of an infectious disease should not be established on the basis of a single test result. A precise diagnosis should take into consideration endemic situation, clinical history, symptomatology, imaging as well as serological data.

In immunocompromised patients and newborns, serological data are of limited value.

References:

Mazzucco, M., Raia, D.D., Marotta, C., Costa, A., Ferrantelli, V., Vitale, F. and Casuccio, A. (2018) Validation of an *Anisakis* sensitization in different population groups and public health impact: A systematic review. Plos one 13.

Kochanowski, M., Gonzalez-Munoz, M., Gomez-Morales, M.A., Gottstein, B., Dabrowska, J., Rozycki, M., Cencek, T., Muller, N. and Boubaker, G. (2019) Comparative analysis of excretory-secretory antigens of *Anisakis simplex*, *Pseudoterranova decipiens* and *Contracaecum osculatum* regarding their applicability for specific serodiagnosis of human anisakidosis based on IgG ELISA. Experimental Parasitology 197, 9-15.



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